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TWO NOVEL PHENOLIC TRITERPENES FROM TRIPTERYGIUM WILFORDII

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Two novel phenolic triterpenes were isolated from *Tripterygium wilfordii* Hook. f., their structures were identified to be 2,3-dihydroxy-1,3,5(10),8-tetra-ene- 6α -(2'-hydroxyethyl)-24-nor-D:A-friedooleanane-29-oic acid 1, named triptotin F, and 2,3-dihydroxy-1,3,5(10),8-tetra-ene- 6β -(2'-hydroxyethyl)-24-nor-D:A-friedooleanane-29-oic acid 2, named triptotin G on the basis of spectroscopic studies.

Keywords: Tripterygium wilfordii Hook. f.; Celastraceae; Triterpenes; Friedooleanane

INTRODUCTION

Tripterygium wilfordii Hook. f., commonly called "Lei Gong Teng" in China, is a perennial twining vine belonging to the family Celastraceae. Tripterygium wilfordii Hook. f. is known to contain a number of constituents, some of which appear to be toxic. Because of its toxicity, "Lei Gong Teng" was only used as pesticide in previous times. Recently, "Lei Gong Teng" and its preparation "Lei Gong Teng Duo Dai" have been used for the treatment of various diseases, including dermatitis, rheumatoid arthritis, systemic acne rosacea, and nephritis [1]. In the course of our continuing search for novel active components from this plant, two novel phenolic triterpenes were isolated and their structures were identified to be 2,3-dihydroxy-1,3,5(10),8-tetra-ene- 6α -(2'-hydroxyethyl)-24-nor-D:

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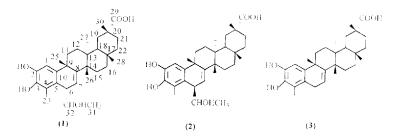


FIGURE 1 Structures of compounds (1)-(3).

A-friedooleanane-29-oic acid 1, named triptotin F, and 2,3-dihydroxy-1,3,5(10),8-tetra-cne- 6β -(2'-hydroxycthyl)-24-nor-D:A-friedooleanane-29-oic acid 2, named triptotin G (Fig. 1).

RESULTS AND DISCUSSION

Compound 1, amorphous powder, high-resolution mass spectrum gave a molecular formula of C₃₁H₄₄O₅. ¹H NMR spectrum revealed the presence of seven methyls [8 1.21, 1.25, 1.44, 1.47, 1.97, 2.50 (each 3H, s), 8 1.61 (3H, d. J = 6.3 Hz)], one methine [δ 4.46 (1H, m)] attached to one oxygen function, one olefinic proton [δ 6.20 (1H, d, J = 6 Hz)], and one aromatic proton [δ 7.13 (1H, s)]. ¹³C NMR spectrum of 1 showed one carbonyl carbon signal (δ 181.3), six aromatic carbon signals [δ 110.7 (d), δ 143.4 (s), δ 145.2 (s), δ 121.4 (s), δ 126.7 (s), δ 142.6 (s)], two olefinic carbon signals [δ 119.6 (d), δ 151.1 (s)], seven methyl carbon signals, seven methylene carbon signals, one oxygenated methine carbon signal [δ 70.8], two methine carbon signals, and five quarternary carbon signals. The formula required ten unsaturated equivalents. Except for one carbonyl group, one benzene ring, and one double bond, the four remaining degrees of unsaturation were ascribed to four carbocyclic systems. Thus triptotin F should be a pentacyclic triterpenoid. We found that the ¹³CNMR spectrum data of 1 were very similar to those of triptohypol C 3 which was isolated from Tripterygium hypoglaucum [2]. Analyses of these signals by HMQC and HMBC experiments proved that the structures of A, C, D, E ring in compound 1 were similar to those of triptohypol C and the double bond was located at C-7 and C-8. From ${}^{1}H - {}^{1}H$ COSY and HMQC spectrum two partial structures (see Fig. 2) were obtained in relation to ring B.

So, the carbon signal at δ 45.4 could be assigned to C-6. In HMBC spectrum (see Fig. 3), the proton signal at δ 1.61 (3H, d, J = 6.3 Hz) was

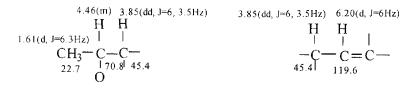


FIGURE 2 Partial structures for ring B of compound 1.

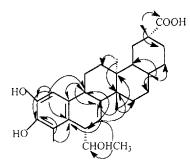


FIGURE 3 HMBC for 1.

correlated with the carbon signals at δ 70.8 (d) and δ 45.4 (d). This fact indicated that CH₃CHOH-was connected with C-6 (δ 45.4). The mass spectrum of 1 showed the fragment ion at m/z 451 due to $[M^+]-C_2H_5O$. The fragment ion at m/z 451 was determined by HRMS to be $C_{29}H_{39}O_4$ (Found: 451.2843, Required: 451.2848). This fact agreed with the structure of 1. Finally, the stereostructure of compound 1 was determined by NOESY (see Fig. 4).

Compound 2, amorphous powder, its ¹³CNMR, IR, MS were similar to those of compound 1. This suggested that the basic carbon skeleton of compound 2 was the same as that of compound 1. ¹HNMR spectrum data of 2 was also similar to that of 1, except for the proton signal at 4.15(1H, t, J = 6.0 Hz, H-6) in case of compound 2 and at 3.85 (1H, dd, J = 6, 3.5 Hz, H-6) in compound 1. This suggested that compound 2 was a stereoisomer of compound 1. Form NOESY spectrum, the proton signal at 4.41(1H, m, H-31) was correlated with the methyl signal at 1.73(3H, s, H₃-25), and this fact clearly showed that — CHOHCH₃ was β at C-6. Thus, the structure of compound 2 was determined as shown.

Up to now, many triterpenes have been isolated from *Celastraceae* plants, *i.e.*, C_{30} type (oleanane, ursane, friedelane), C_{29} type (pristimerin, celastrol), C_{28} type (tingenone, iguesterin), but C_{31} type triterpenes from *Celastraceae* plants have never been described. Compound 1 and 2 which belonged to C-31 type were first isolated from *Celastraceae* plants.

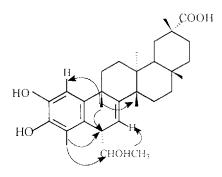


FIGURE 4 NOESY for 1.

EXPERIMENTAL SECTION

General Experimental Procedures

IR spectra were recorded on a Perkin-Elmer 599B IR spectrometer. Optical rotations were measured on a JASCO DIP-181 polarimeter, using a 10-cm microcell. MS spectra were measured on a MAT-711 spectrometer, ¹H-NMR and ¹³C-NMR spectra were recorded on Bruker AM-400 instrument with TMS as internal standard. Silica gel chromatography was performed using silica gel 60H at Qingdao Haiyang Chemical Group Co.

Plant Material

The roots of *Tripterygium wilfordii* were collected in Fujian Province, China. The plant material was identified by Prof. Guan-Yuan Gu, Scientific and Technical Archives of Shanghai Medical University, Shanghai, China.

Extraction and Isolation

The air-dried roots (200 kg) of *Tripterygium wilfordii* were powdered and extracted with 95% EtOH. The EtOH extract was extracted with CHCl₃. The CHCl₃-soluble fraction (500 g) was chromatographed on silica gel with CHCl₃--MeOH (95:5, 9:1, 8:2, MeOH) to give **5** fractions (A-E). Fraction **B** (95g) was chromatographed over a silica gel column with CHCl₃--MeOH (98:2, 95:5, 9:1, MeOH) to give **8** fractions. Fraction **4** (15g) was chromatographed over a silica gel column eluting with CH₂Cl₂--MeOH (98:2) to give **3** fractions (fractions **4.1-4.3**), and frac-

tion 4.1 (7.5 g) was then subjected to column chromatography on silica gel with cyclohexane – acetone (5:1, 3:1, 1:1) to give 4 fractions (fractions 4.1.1–4.1.4). Fraction 4.1.3 (2.5 g) was chromatographed over a silica gel column eluting with CH_2Cl_2 –MeOH (98:2) to afford 3 fractions (fractions 4.1.3.1–4.1.3.3). Fraction 4.1.3.3 (1.1 g) was subjected to RP-18 column chromatography with MeOH–H₂O (3:2) and column chromatography on silica gel with CH_2Cl_2 –MeOH (98:2) to give compound 1 (37.6 mg). Fraction 6 (25 g) was chromatographed over a silica gel column eluting with petroleum cher-ethyl acetate (5:1, 5:2, 3:1, 1:1) to give 9 fractions (fractions 6.1–6.9). Fraction 6.5 (3.7 g) was chromatographed over a silica gel column eluting with CH_2Cl_2 –MeOH (98:2) to give compound 2 (250 mg).

Triptotin **F** (1), amorphous powder; $[\alpha]_D^{20} - 67.4$ (*c* 0.38, CHCl₃); IR $\nu_{\text{max}}^{\text{KBr}} \text{ cm}^{-1}$: 3400, 1697, 1606, 1461, 1209, 1202, 871; ¹HNMR (C₅D₅N, 400 MHz): δ 1.21 (3H, s, H₃-28), 1.25 (3H, s, H₃-27), 1.44 (3H, s, H₃-26), 1.47 (3H, s, H₃-30), 1.97 (3H, s, H₃-25), 2.50 (3H, s, H₃-23), 1.61 (3H, d, J = 6.3 Hz, H₃-31), 3.85 (1H, dd, J = 6, 3.5 Hz, H-6), 4.46 (1H, m, H-32), 6.20 (1H, d, J = 6 Hz, H-7), 7.13 (1H, s, H-1), ¹³CNMR (see Tab. I); EIMS m/z (rel.int.): 496[M]⁺ (3), 451 (19), 215 (21), 201 (100), 95 (17), 69 (10), HRMS m/z 496.3155 [M]⁺ (C₃₁H₄₄O₅, required 496.3189).

Triptotin G (2), amorphous powder; $[\alpha]_D^{20} - 56.0$ (*c* 0.28, MeOH); IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3412, 1700, 1462, 1377, 1288, 1207, 1018, 871; ¹HNMR (C₅D₅N, 400 MHz): δ 1.20 (3H, s, H₃-28), 1.23 (3H, s, H₃-27), 1.37 (3H, s, H₃-26), 1.46 (3H, s, H₃-30), 1.73 (3H, s, H₃-25), 2.75 (3H, s, H₃-23), 1.45 (3H,

С	1	2	3	С	1	2	3
1	110.7	110.6	109.8	17	31.1	30.9	30.9
2	143.4	141.3	140.7	18	45.1	44.9	44.8
3	145.2	143.1	142.7	19	31.3	31.1	31.2
4	121.4	122.5	121.4	20	40.8	40.6	40.6
5	126.7	126.0	124.2	21	31.4	31.2	30.5
6	45.4	46.1	28.3	22	35.9	35.7	35.5
7	119.6	120.9	118.4	23	13.4	13.8	12.2
8	151.1	+	149.9	24			
9	38.3	38.1	37.1	25	36.5	35.7	34.8
10	142.6	145.1	145.1	26	22.7	22.4	23.1
11	37.6	37.7	35.1	27	18.9	18.8	18.8
12	31.4	30.6	30.7	28	32.0	31.9	31.8
13	38.8	38.7	38.2	29	181.3	181.0	181.1
14	44.7	44.5	44.0	30	33.5	33.3	33.3
15	29.9	29.7	29.4	31	22.7	21.4	
16	37.6	37.4	37.3	32	70.8	72.1	

TABLE I ¹³CNMR data of compound $1-3^*$

* Compound 1-3 in $C_5D_5N_2$

⁺ Signal is hidden by a solvent peak.

d. J = 6 Hz, H₃-31), 4.15 (1H, t, J = 6 Hz, H-6), 4.41 (1H, m, H-32), 6.27 (1H, d, J = 6 Hz, H-7), 7.24 (1H, s, H-1); ¹³CNMR (see Tab. 1); EIMS m/z (rel.int.): 496[M]⁺ (3), 451 (16), 215 (20), 201 (100); HRMS m/z 496.3193 [M]⁺ (C₃₁H₄₄O₅, required 496.3189).

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